

Geometrical Origins of Contractility in Disordered Actomyosin Networks

Martin Lenz*

Université Paris-Sud, CNRS, LPTMS, UMR 8626, Orsay 91405, France

(Received 14 April 2014; published 2 October 2014)

Movement within eukaryotic cells largely originates from localized forces exerted by myosin motors on scaffolds of actin filaments. Although individual motors locally exert both contractile and extensile forces, large actomyosin structures at the cellular scale are overwhelmingly contractile, suggesting that the scaffold serves to favor contraction over extension. While this mechanism is well understood in highly organized striated muscle, its origin in disordered networks such as the cell cortex is unknown. Here, we develop a mathematical model of the actin scaffold's local two- or three-dimensional mechanics and identify four competing contraction mechanisms. We predict that one mechanism dominates, whereby local deformations of the actin break the balance between contraction and extension. In this mechanism, contractile forces result mostly from motors plucking the filaments transversely rather than buckling them longitudinally. These findings shed light on recent *in vitro* experiments and provide a new geometrical understanding of contractility in the myriad of disordered actomyosin systems found *in vivo*.

DOI: 10.1103/PhysRevX.4.041002

Subject Areas: Biological Physics, Complex Systems

I. INTRODUCTION

The structure and motion of living cells is largely controlled by the continuous remodeling of their cytoskeleton, which crucially involves the contractility of networks of actin filaments (F-actin) and myosin molecular motors. How macroscopic motion emerges from the protein-scale interactions between these components was first understood in the context of striated muscle [1]. There, individual myosins are assembled into so-called “thick filaments,” bottlebrush-shaped clusters of myosin capable of binding several actin filaments and sliding along them for long distances—for brevity, we refer to them as “motors” in the following. In striated muscle, F-actin and motors are strongly organized into a periodic array of so-called sarcomeres, contractile units where the sliding action of the motors is harnessed to produce contraction through F-actin's geometrical arrangement [Fig. 1(a)].

However, in many biological situations, contractile F-actin and myosin assemblies—be they one-dimensional bundles or two- or three-dimensional networks—lack the organization found in sarcomeres [2–8]. While the biochemical processes inducing the relative motion of the motors and filaments are similar to the ones involved in striated muscle, here the geometrical mechanisms used to convert this relative motion into contraction in the absence of organization are less clear. Indeed, the filaments and motors do not have an intrinsic propensity towards

contraction, and can *a priori* yield extension just as easily. Figure 1(b) illustrates this property in a simple one-dimensional example. Most theoretical models of disordered actomyosin contractility circumvent this

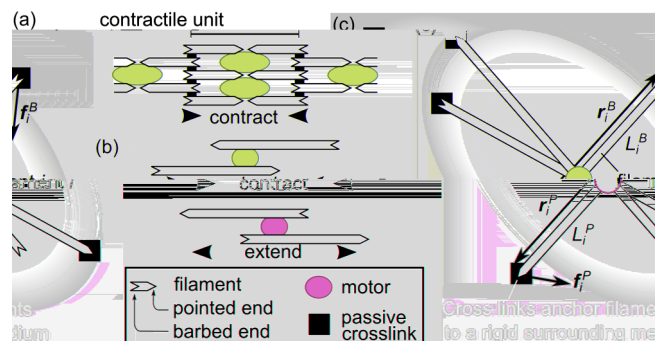


FIG. 1. Geometrical foundations of contractility. Motors bound to filaments slide toward their “barbed ends,” as for myosin II thick filaments. (a) In striated muscle, motors are localized close to the filaments’ pointed ends. When activated, every motor pulls in the neighboring filaments and thus induces local contraction. (b) If filament polarities are not carefully selected, striated muscle-like locally contractile configurations (top) are just as likely as extensile ones (bottom), and the overall behavior of the actomyosin assembly is unclear. (c) The symmetry between contraction and extension subsists in a two- or three-dimensional network. Throughout this article, filament extremities may or may not be cross-linked to the surrounding medium. Even though this is not represented here, cross-linked filaments extend beyond the cross-links and farther into this medium, and thus cannot freely rotate around these cross-links. Thus, the barbed and pointed arrow ends on this schematic merely indicate filament polarity; the actual barbed and pointed ends of cross-linked filaments are typically farther away, inside the surrounding medium.

*martin.lenz@u-psud.fr

Published by the American Physical Society under the terms of the Creative Commons Attribution 3.0 License. Further distribution of this work must maintain attribution to the author(s) and the published article's title, journal citation, and DOI.

question by assuming from the onset that motors either induce an average contractile stress in the actomyosin medium [9] or, in more detailed descriptions, that they give rise to localized contractile force dipoles [10]. These studies then typically move on to consider the macroscopic consequences of such mesoscopic behaviors. In contrast, in this paper, we adopt a different focus and ask how the contractility emerges from the networks' microscopic components in the first place.

This question is most easily discussed in one-dimensional actomyosin assemblies, i.e., actomyosin bundles. There, *in vitro* experiments demonstrate that sarcomerelike organization is not necessary for contraction [11], and thus, the symmetry between contraction and extension illustrated in Fig. 1(b) is spontaneously broken. Because geometry in one dimension is very simple, there are strong geometrical constraints on the type of mechanisms that can lead to such symmetry breaking [12]. Combining these theoretical constraints with further experiments, we have recently shown that F-actin buckling under longitudinal compression enables contraction by favoring local filament collapse in the absence of sarcomerelike organization [13].

The situation in two- and three-dimensional actomyosin networks is more complex than that of bundles. There, too, contraction arises in random-polarity, disordered *in vitro* networks [14–16]. From a theoretical standpoint, however, geometry in two or three dimensions is considerably richer than in one dimension. As a result, several mechanisms can *a priori* give rise to contraction, and symmetry considerations are less easily exploited than in bundles. Accordingly, a range of mechanisms for the emergence of actomyosin contraction have previously been invoked in different levels of detail, ranging from cartoon pictures [10,17] to more quantitative numerical [18] and analytical [19] approaches. However, there is no consensus regarding their relative roles in either *in vivo* or *in vitro* actomyosin contractility.

Here, we present the first comprehensive comparison of contractility-inducing mechanisms in disordered cytoskeletal networks. We first exploit symmetry considerations in two and three dimensions to identify all possible local contraction mechanisms. We then study them individually and compare their relative magnitudes, thus determining the dominant cause of contractility as a function of experimental conditions. Filament deformation is found to play a crucial role in most relevant regimes.

II. REQUIREMENTS FOR CONTRACTION

We first show that, unlike in striated muscle, filament sliding alone is not sufficient to induce contraction in disordered networks. We do this by studying a minimal, sliding-only model and demonstrating that it cannot yield contractility.

We consider a single motor bound to multiple filaments. The filaments are themselves cross-linked to a surrounding rigid external medium as illustrated in Fig. 1(c). We show that overall network contraction cannot occur under the following main assumptions: 1. the motor stall force does not depend on its position, 2. the motor is pointlike, 3. the motor is undeformable, 4. filaments behave as rigid rods. The essence of our argument is as follows. In a network, individual motors may exert either contractile or extensile local forces depending on the polarities of the neighboring filaments [as in Fig. 1(b)]. In a disordered system satisfying the above assumptions, there are as many contractile as extensile motors and the forces produced by the former exactly compensate those produced by the latter. Therefore, the network does not contract *overall*. Thus, overall disordered actomyosin contractility requires the breaking of at least one of these assumptions.

We first introduce some notation. The overall contractility of a rigid disordered network is characterized by the average local force dipole [20] \mathcal{D} exerted by an individual motor, where

$$\mathcal{D} = \sum_i \sum_{a=B,P} \mathbf{r}_i^a \cdot \mathbf{f}_i^a. \quad (1)$$

Here, i indexes the filaments as in Fig. 1(c), $a = B, P$ denote the directions of the filaments' barbed and pointed ends respectively; therefore, each term of the double sum over i and a corresponds to a filament section in contact with the motor. For instance, for the example of Fig. 1(c), $i \in \{1, 2, 3\}$, and thus the sum has six terms. The position vector of a cross-link is denoted as \mathbf{r}_i^a , and \mathbf{f}_i^a is the force exerted on it by filament i [Fig. 1(c)]. A negative (positive) \mathcal{D} denotes a contractile (extensile) system. The portion of filament between the motor and cross-linker (i, a) is referred to as a "filament section," and we denote its length by L_i^a . At steady state, the motor exerts a longitudinal "stall force" f directed toward the pointed end of each filament. This force is transmitted to the cross-linkers through the stretching and compression of the rigid filaments. We thus introduce the stretching moduli $k(L_i^a)$ of the filament sections, i.e., their longitudinal Hookean spring constants. In general, \mathcal{D} is a function of f , the L_i^a 's, and the $k(L_i^a)$'s.

We now present our argument in more detail. Consider the filament-motor system of Fig. 1(c). For rigid filaments, linear elasticity applies and the forces \mathbf{f}_i^a exerted on the cross-linkers are proportional to the motor's stall force. Using Eq. (1) and noting that the \mathbf{r}_i^a are constants due to the rigidity of the external medium, this implies

$$\mathcal{D} \propto f. \quad (2)$$

Now, consider a new system obtained by reversing the filament polarities of the original system—i.e., exchanging the barbed and pointed ends in Fig. 1(c). As polarities are reversed, the motor reverses its sliding direction on each

filament, which is equivalent to changing the sign of its stall force: $f^{\text{reversed}} = -f$. Using Eq. (2), the polarity-reversed force dipole is thus $\mathcal{D}^{\text{reversed}} = -\mathcal{D}$. Hence, if the original system generates contractile forces, then the polarity-reversed system generates the same amount of extensile forces.

To complete our reasoning, we consider a large-scale disordered network comprising many filament-motor systems embedded in a rigid medium. The rigid medium can be described as linearly elastic, and thus the network's overall contractile dipole is proportional to the average dipole of a filament-motor system. Because of the network's disorder, any individual filament-motor system is just as likely to occur as its polarity-reversed counterpart. Averaging the force dipoles over the whole network, we thus find that individual contractile and extensile dipoles cancel mutually. From this, we conclude that the network has an overall vanishing contractile force dipole, which completes our proof.

This result is quite general, as it requires only a minimal form of disorder, namely, polarity-reversal symmetry (i.e., the property that any arrangement of filaments is just as likely as its polarity-reversed counterpart). This is a variant of a more powerful argument valid for one-dimensional bundles [12]; a more formal presentation is given in the Supplemental Material [21]. Interestingly, this polarity-reversal symmetry can be broken not only through sarcomeric organization, which yields contractility, but also in solution through a dynamical process of motor-filament coalescence and sliding, which favors extension [22]. However, this process is not relevant for the rigid networks considered here.

III. COMPETING CONTRACTILITY MECHANISMS

While the model considered in the previous section cannot generate contractility, such contractility is experimentally observed in actomyosin networks [11,14,16,23]. This discrepancy implies that this model is an oversimplification: one or several of its assumptions must be violated. By successively relaxing each of these assumptions, here we systematically review all essential contraction mechanisms and predict the magnitude of the associated contractile forces.

A. Position-dependent stall force

Early models of nonsarcomeric actomyosin bundles [24,25] and networks [19] proposed that motors stop upon reaching the filament barbed ends, staying there for some time before eventually detaching. Although experimental evidence for this behavior in actomyosin is lacking, the resulting accumulation of immobile motors at the filament barbed ends would generate sarcomerelike cross-linking [Fig. 1(a)] and thus favor contraction.

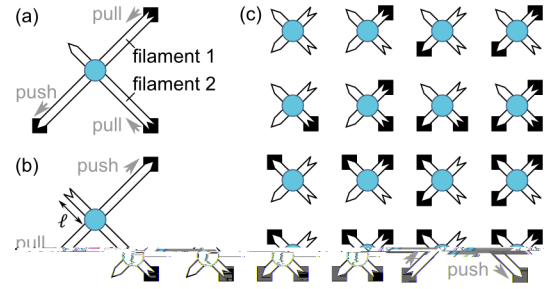


FIG. 2. Contraction induced by a position-dependent stall force. As in Fig. 1 in all other figures, black squares and blue circles represent cross-links and motors, respectively. (a) Motors in the vicinity of a pointed end typically induce an overall contractile (pulling) force dipole as indicated by gray arrows representing the projection of the forces on the direction of the filaments. (b) Motors close to a barbed end have the opposite effect. (c) We characterize the resulting net contractility by averaging over all possible local cross-linking configurations.

We consider a two-filament system where the motor operation has such a dependence on its distance ℓ from the barbed end [Figs. 2(a) and 2(b)]. Specifically, we assume that the stall force exerted on a filament vanishes [26] as the motor approaches its barbed end closer than a distance $d \ll \xi$:

$$f(\ell) = f(1 - e^{-\ell/d}). \quad (3)$$

The force dipole exerted by a specific configuration depends on whether each of its filament ends is cross-linked to the surrounding medium. For instance, we compute the force dipole associated with Fig. 2(a) by resolving force balance under the assumption that the passive cross-links impose clamped boundary conditions:

$$\mathcal{D} = -f(\ell_2)L_2^B - f(\ell_1)\frac{L_1^B k(L_1^B) - L_1^P k(L_1^P)}{k(L_1^P) + k(L_1^B)}, \quad (4)$$

where ℓ_1 and ℓ_2 are the distances from the motor to the barbed ends of filaments 1 and 2, respectively. The first term on the right-hand side of Eq. (4) is always negative, indicating that filament 2 transmits the stall force $f(\ell_2)$ to the bottom right-hand cross-link, exerting only pulling forces. In contrast, the second term can be either positive or negative as filament 1 distributes this force across two cross-links and thus exerts both pulling and pushing force. Note that Eq. (4) is derived in the rigid filament limit $\epsilon = f/\xi k(\xi) \rightarrow 0$, where ξ is the average distance between the motor and the neighboring cross-linker.

Similar to our derivation of Eq. (4), we compute the expressions of the force dipoles associated with each possible motor-cross-linker configuration [Fig. 2(c)]. Assuming that both the motor and the cross-linkers are uniformly distributed on the filaments, we use these expressions to compute the force dipole averaged over

all possible configurations and over filament section lengths:

$$\langle \mathcal{D}_{\text{dwell}} \rangle_{d \ll \xi \ll L_f} \sim -\frac{2d}{L_f} f \xi, \quad (5)$$

where L_f is the total length of a filament. The condition $L_f \gg \xi$ guarantees that filaments are cross-linked several times and, therefore, are not free to rotate.

To understand why the dipole of Eq. (5) is contractile, we remind ourselves that if the stall force were the same irrespective of motor position, the contractile force dipole of Fig. 2(a) would exactly cancel the extensile dipole of its polarity-reversed image [Fig. 2(b)]. According to Eq. (3), however, the motor in Fig. 2(b) exerts a weaker force on filament 2 than in Fig. 2(a) due to the proximity of the filament barbed end. The contractility of Fig. 2(a) thus exceeds the extensility of Fig. 2(b), resulting in overall contractility. The corresponding average force dipole [Eq. (5)] is thus proportional to the probability d/L_f for the motor to be within a distance d of a barbed end, multiplied by the typical force dipole $f\xi$.

B. Finite motor size

Unlike the pointlike motors considered above, a finite-size motor bound to two filaments is not constrained to remain at their intersection. It tends to move towards their barbed ends, as shown in Fig. 3(a). This motion breaks the equivalence between barbed and pointed end (also known as polarity-reversal symmetry), thus enabling contraction [18].

We consider two filaments intersecting at an angle θ as in Fig. 3(a). All filament sections are cross-linked, have length ξ , and are considered rigid. The motor is modeled as a rigid dumbbell of length L_m whose heads slide on the filaments until their stall force is reached. To enforce this condition, we minimize the pseudoenergy [18]

$$E_m = -f(L_1^P + L_2^P) \quad (6)$$

under the constraint of constant L_m . Once the motor is stalled, the midpoint of the motor is offset from the filament intersection by a distance $L_m/[2 \tan(\theta/2)]$. Computing the force dipole $\mathcal{D}(\theta)$ from force balance as in the previous section, we find that small values of θ yield large motor displacements and thus large force dipoles. We average this force dipole over angles in three dimensions using $k(L) \propto L^{-4}$, as expected for filaments with predominantly entropic elasticity [27,28]:

$$\langle \mathcal{D}_{\text{finite size}} \rangle = \frac{1}{2} \int_0^\pi \mathcal{D}_{\text{finite size}}(\theta) \sin \theta d\theta \Big|_{L_m \ll \xi} \sim -16fL_m. \quad (7)$$

To understand the source of this contractile dipole, we draw an analogy between the motor and the slider of a

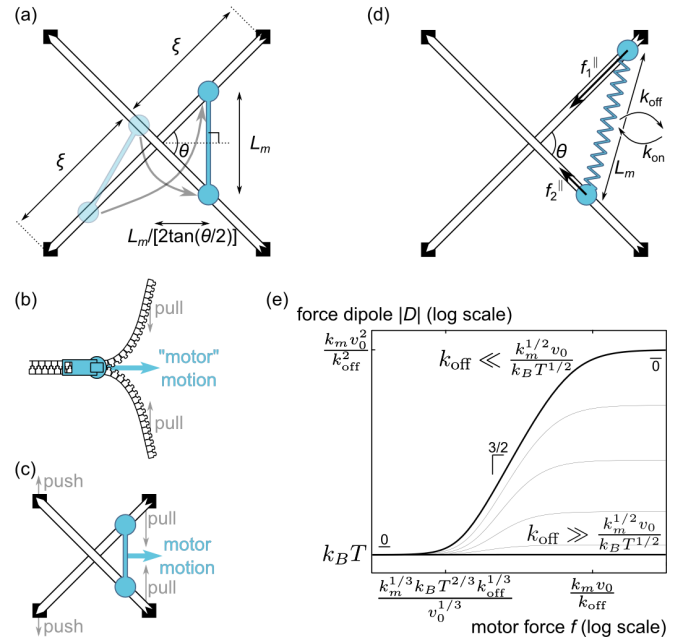


FIG. 3. Contraction induced by finite-size and deformable motors. (a) A finite-size motor minimizes the pseudoenergy [Eq. (6)] by orienting itself perpendicular to the bisector of the filaments (dotted line) as shown by the gray arrows. (b) The contractility induced by such a motor is analogous to the closing force (thin gray arrows) of a zipper when its slider is being slid shut (thick cyan arrow). (c) In practice, the zipperlike pulling forces exerted at the barbed end cross-links are partially compensated by pointed end pushing forces. (d) An attaching-detaching flexible motor generates contractility in a similar fashion. (e) Scaling regimes for the deformable motor dipole [Eq. (10)]. Black lines present the limits of small (top curve) and large (bottom curve) detachment rate k_{off} and thin gray lines display intermediate regimes.

zipper [Figs. 3(b) and 3(c)]. Assimilating the motor's propensity to slide along the filaments to a closing force applied on the zipper tab, we see that the motor pulls the filament barbed ends together as it progresses, just like the two sides of the zipper chain are pulled together as the zipper closes. This induces a predominantly contractile force dipole.

Importantly, this zipper effect induces contraction only if the motor is displaced from the intersection of the filaments, as is the case for a finite-size motor. Indeed, while the motor pulls on the filaments' barbed end cross-links, it also pushes out on the pointed end cross-links, as shown in Fig. 3(c). These two effects compensate exactly for vanishing motor length $L_m = 0$, suggesting that for small L_m , \mathcal{D} is generically proportional to L_m . Additionally, \mathcal{D} is proportional to f in the rigid filament limit as discussed above. We thus expect zipperlike contractility to scale as

$$\mathcal{D} \approx -fL_m, \quad (8)$$

consistent with the result of Eq. (7).

C. Deformable motor

We now consider a variant of the previous model where an initially pointlike motor can be stretched to a nonzero size, again implying zipperlike contractility. We also consider motor attachment and detachment, as experiments indicate that it can have a significant influence on force buildup in the regimes where the present mechanism will eventually be found to dominate [13].

We consider the geometry of Fig. 3(d) with a motor of variable length L_m and an associated stretching energy $E_s = k_m L_m^2/2$, where k_m plays the role of a motor “spring constant.” The motor detaches from the filaments at a fixed rate k_{off} and reattaches with $k_{\text{on}} = k_{\text{on}}^0 \exp(-E_s/k_B T)$, thus satisfying detailed balance. This rate is substantial only in the region where $E_s \approx k_B T$, implying a motor length $L_m \approx \sqrt{k_B T/k_m}$ of the order of a detached motor’s root-mean-square thermal extension. We define the ratio $\eta = \sqrt{k_B T/k_m}/\xi$ of typical motor size to filament section length and consider the stiff motor limit $\eta \ll 1$, analogous to the $L_m/\xi \ll 1$ regime considered above. The velocity v_i of motor head i depends on the projection f_i^{\parallel} of the motor tension onto the direction of the filament through its force-velocity relationship, assumed linear for simplicity:

$$v_i = v_0(1 - f_i^{\parallel}/f), \quad (9)$$

where v_0 is the motor’s unloaded velocity. Taking into account the stochastic attachment and detachment of the motor and its sliding under thermal agitation, we calculate the probability to find it in a given position on the filaments and average the resulting steady-state force dipole over all angles θ in three dimensions (see Supplemental Material [21]). We find

$$\langle \mathcal{D}_{\text{ext}} \rangle = -8\pi k_B T \left[1 + \beta^2 \frac{\sqrt{2+\alpha} - \sqrt{1+\alpha}}{\sqrt{\alpha(1+\alpha)}(2+\alpha)} \right], \quad (10)$$

where $\alpha = k_{\text{off}} f/2v_0 k_m$ is the ratio of the time required to reach stall to the spontaneous detachment time and $\beta = f/\sqrt{k_m k_B T}$ is the ratio of the motor stall force to the force scale over which the attachment rate varies. The two terms in the square brackets of Eq. (10) correspond to two different origins for contractility. We denote the first, β -independent term as $\mathcal{D}_{\text{ext}}^{\text{passive}}$. This term does not involve the motor stall force and describes the equilibrium effects of motor binding, which tends to pull the filaments together and exert a contractile force dipole:

$$\mathcal{D}_{\text{ext}}^{\text{passive}} \approx -k_B T. \quad (11)$$

The second term, denoted here by $\mathcal{D}_{\text{ext}}^{\text{active}}$, has two distinct asymptotic regimes. If $\alpha \gg 1$, the motor spontaneously detaches long before reaching stall, yielding a typical

extension $L_m \approx v_0/k_{\text{off}}$. In this regime, the motor exerts a typical force $\approx k_m L_m$ on the filaments, equal to the tension of the spring. The resulting typical force dipole is given by Eq. (8) as

$$\mathcal{D}_{\text{ext}}^{\text{active}} \underset{\alpha \gg 1}{\approx} -(k_m L_m) L_m \approx -k_m v_0^2/k_{\text{off}}^2. \quad (12)$$

Conversely, if $\alpha \ll 1$, the motor reaches stall for moderate angles, implying a force f and an extension $L_m = f/k_m$. However, in this case, the average force dipole is not dominated by moderate angles, but rather by small angle configurations for which $\theta \approx \sqrt{\alpha}$. In these configurations, the two filaments are so close to parallel that the motor can slide without stalling until its spontaneous detachment. Similar to the typical motor of the $\alpha \gg 1$ regime, these motors have $L_m \approx v_0 \theta/k_{\text{off}}$ and a spring force $\approx k_m L_m$. In the regime $\theta \approx \sqrt{\alpha}$, this yields a force dipole $\mathcal{D}_{\text{ext}}^{\text{active}}(\theta \approx \sqrt{\alpha}) \approx -k_m v_0^2 \alpha/k_{\text{off}}^2$. Taking into account the fact that motors can bind to both filaments only in the region where these filaments are within a distance $\approx L_m$ of each other, and noting that this region is much larger for pair of filaments separated by a small θ , we find that motors in the small-angle regime $\theta \approx \sqrt{\alpha} \ll 1$ represent a fraction $\sqrt{\alpha}$ of the total motor population. This leads to an average force dipole

$$\mathcal{D}_{\text{ext}}^{\text{active}} \underset{\alpha \ll 1}{\approx} \sqrt{\alpha} \mathcal{D}_{\text{ext}}^{\text{active}}(\theta \approx \sqrt{\alpha}) \approx -\frac{f^{3/2} v_0^{1/2}}{k_m^{1/2} k_{\text{off}}^{1/2}}. \quad (13)$$

As in the previous section, configurations where the filaments are nearly parallel exert disproportionately large force dipoles that dominate the average.

Figure 3(e) ties together the asymptotic regimes discussed here as a function of the original model parameters. In the large detachment rate regime (bottom black curve), detachment is too fast to allow the motors to escape their initial binding region and the force dipole is dominated by its passive component. Conversely, if detachment is slow (top black curve), the magnitude of the motor’s stall force matters. The passive dipole still prevails for small forces, while intermediate and large forces are, respectively, dominated by the active regimes of Eqs. (13) and (12).

D. Deformable filaments

While the previous sections assumed straight, stiff filaments, here we consider the effect of filament deformation on contractility. Related mechanisms were previously discussed for actomyosin bundles [12,13,29] and gels [10,17]. We discuss two asymptotic regimes: small motor forces, which mostly induce filament bending, and large motor forces, which significantly stretch out the filaments’ thermal fluctuations. The typical force separating the two regimes is $f \approx k_B T \ell_p^{1/2}/\xi^{3/2}$, i.e., the transverse force required to pull out a significant fraction of these fluctuations.

1. Small-force regime $f \ll k_B T \ell_p^{1/2} / \xi^{3/2}$

In the absence of significant filament stretching, we consider the filament profile as a weakly perturbed straight line described by the wormlike chain model [Fig. 4(a)]:

$$E = 2 \left[\frac{k_B T \ell_p}{2} \int_{-\xi}^{\xi} \left(\frac{d^2 x}{dz^2} \right)^2 dz - f \delta \ell \right], \quad (14)$$

where z is the filament's longitudinal direction, x its transverse displacement, $\delta \ell$ the motor's longitudinal displacement, and ℓ_p the filament persistence length. The last term of Eq. (14) represents the motor pseudoenergy as in Eq. (6), and contact of the motor with the filaments imposes $x(\delta \ell) = \delta \ell \tan(\theta/2)$.

In this problem, the motor can only progress toward the barbed ends by deforming the filaments. The amplitude x of this deformation is obtained by balancing the filament and motor forces, implying that the filament and motor (pseudo)energies are of comparable magnitudes and so that $x \approx f \xi^3 / (k_B T \ell_p)$. The dominant source of contractile forces is different from the zipperlike mechanism discussed above. Here, the displacement of the motor plucks the filament like the finger of the musician does the string of a harp; interestingly, this mode of deformation induces much larger contractile force than filament buckling [10,17] in the $\xi \ll \ell_p$ limit. A small transverse displacement $\approx x$ induces a longitudinal strain $\gamma \approx (x/\xi)^2$ along the filament. This in turn implies a filament tension $T \approx (k_B T \ell_p^2 / \xi^4) \gamma$, where $k_B T \ell_p^2 / \xi^4$ is the typical entropic stretching modulus of the filament [27]. The resulting force dipole scales as $\mathcal{D} \approx -T \xi \approx -f^2 \xi^2 / k_B T$. A detailed calculation (see Supplemental Material [21]) reveals that small angles again have a disproportionately large contribution to the average force dipole, adding a (weak) logarithmic correction to the predicted scaling:

$$\langle \mathcal{D}_{\text{bend}} \rangle_{f \ll k_B T \ell_p^{1/2} / \xi^{3/2}, \xi \ll \ell_p} \sim -\frac{3}{16} \frac{f^2 \xi^2}{k_B T} \ln \left(\frac{k_B T \ell_p^{1/2}}{c_{\text{bend}} f \xi^{3/2}} \right), \quad (15)$$

where $c_{\text{bend}} \approx 0.191859$. This expression holds until the thermal fluctuations of the filament, which are responsible for its elongational compliance, are pulled out. This occurs for $\gamma \approx \xi / \ell_p$, implying that the small-force regime discussed here is defined by $f \ll k_B T \ell_p^{1/2} / \xi^{3/2}$, as indicated in Eq. (15).

2. Large-force regime $f \gg k_B T \ell_p^{1/2} / \xi^{3/2}$

Under strong extension, the entropic fluctuations of the semiflexible filaments are entirely pulled out, freeing an excess length $s \approx \xi^2 / \ell_p \ll \xi$, as shown in Fig. 4(b). The filaments are, therefore, analogous to inextensible strings of fixed arclength $2\xi + s$, implying a transverse displacement $x \approx \sqrt{\xi s}$. Since the stalled motor exerts a transverse force f , force balance along the x direction imposes a longitudinal filament tension $T \approx f \xi / x$. The force dipole is thus essentially equal to $T \xi \approx f \sqrt{\xi \ell_p}$, consistent with the result of a detailed calculation (see Supplemental Material [21]):

$$\langle \mathcal{D}_{\text{stretch}} \rangle_{f \gg k_B T \ell_p^{1/2} / \xi^{3/2}, \xi \ll \ell_p} \sim c_{\text{stretch}} f \sqrt{\xi \ell_p}, \quad (16)$$

with a numerical prefactor $c_{\text{stretch}} \approx 1.73463$.

We illustrate the crossover between the small- and large-force regimes in Fig. 4(c).

IV. RELATIVE IMPORTANCE OF EACH MECHANISM

To determine the dominant contraction mechanism, we compare the force dipoles induced by each mechanism presented above as a function of two experimentally controllable parameters: the number of myosin heads per myosin thick filament N [30] and the inter-cross-link length ξ . We consider actin filaments with length $L_f = 5 \mu\text{m}$ and persistence length $\ell_p = 10 \mu\text{m}$. The myosin thick filaments have length $L_m = N l_m$ with $l_m = 3 \text{ nm}$, unloaded velocity $v_0 = 200 \text{ nm s}^{-1}$, and stall force $f = N f_0$. Since motor heads spend only a fraction of their time bound to actin, we estimate $f_0 = 0.1 \text{ pN}$ on average. We use $k_m = \mu / L_m$ with $\mu = 45 \text{ nN}$ a typical protein filament rigidity [31]. Myosin II has a duty ratio $1 - p_d \approx 4\%$ and a characteristic attachment-detachment time of $\tau_d = 3 \text{ ms}$ [32], yielding a motor detachment rate $k_{\text{off}} = p_d^N / \tau_d$. Finally, we assume that motors slow down when their distance to the barbed end is comparable to their size: $d = L_m$.

The colored domains in Fig. 5 indicate as a function of N and ξ which of the four dipoles computed in Sec. III has the largest magnitude [Eqs. (5), (7), (10), (15)–(16)]. The bottom right-hand half of the diagram is left blank, as it involves very large motors ($L_m > \xi$) not captured by our

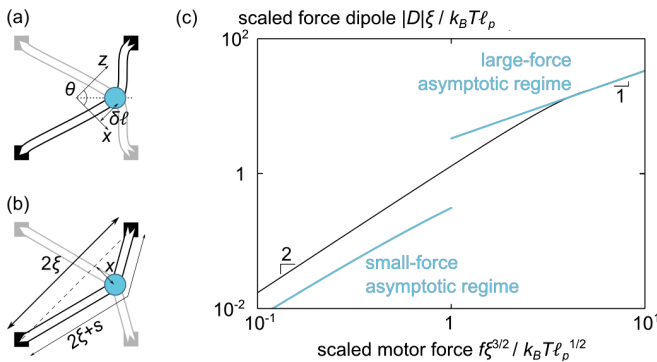


FIG. 4. Contraction induced by filament deformation. (a) For small motor forces, the cost of filament deformation is mainly due to bending. The (x, z) coordinate system is given for the darker filament. (b) For large motor forces, filaments are fully stretched. (c) Crossover of the force dipole \mathcal{D} between the asymptotic regimes of Eqs. (15) and (16). The interpolating black line is discussed in the Supplemental Material [21].

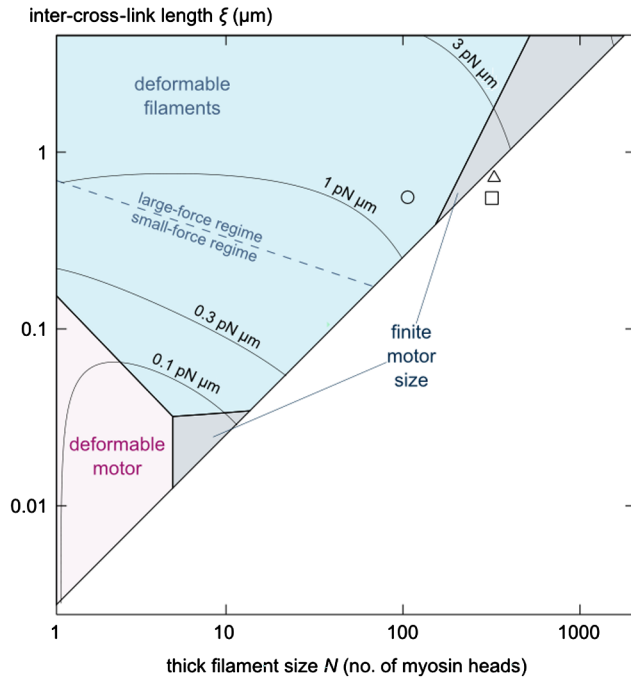


FIG. 5. Contractile forces as a function of experimentally controllable parameters. Colors identify the dominant contraction mechanism in each parameter regime. Contours indicate the magnitude of the contractile force dipole per myosin head $\langle \mathcal{D} \rangle / N$. Symbols indicate the *in vitro* experimental regimes of Ref. [16] (circle), Refs. [15,33] (square) and Ref. [34] (triangle).

current approach; our assumptions $\xi < \ell_p$ and $d < \xi < L_f$ are satisfied throughout the top left-hand (colored) half. The finite motor size mechanism tends to dominate in the vicinity of the diagonal where the motor size L_m is largest. The deformable motor mechanism dominates in the bottom left-hand corner of the diagram; for these small values of N and ξ , and given that the myosin thick filaments are hardly stretchable ($\mu \gg f$), thermal agitation dominates and $\mathcal{D}_{\text{ext}}^{\text{active}} \ll \mathcal{D}_{\text{ext}}^{\text{passive}}$. Deformable filament mechanisms govern contractility in large- ξ regions where the filament sections are most flexible and can thus be deformed by motor forces. Finally, the position-dependent stall force mechanism is always negligible in front of the finite-size motor mechanism; thus, it never dominates contractility. This picture is remarkably insensitive on precise parameter values (see Supplemental Material [21]).

We next consider the total force dipole $\langle \mathcal{D} \rangle$, defined as the sum of the four force dipoles computed in Sec. III. The magnitude of the total dipole per myosin head $\langle \mathcal{D} \rangle / N$ is represented by contour lines in Fig. 5. In the $\xi \gtrsim 0.3 \mu\text{m}$ region, these forces compare with the force dipole exerted by a myosin head in striated muscle $\mathcal{D}/N = (500 \text{ pN} \times 3 \mu\text{m})/300 = 5 \text{ pN} \mu\text{m}$; filament deformation-based mechanisms dominate most of this parameter region. Conversely, for $\xi \lesssim 0.3 \mu\text{m}$, forces are much weaker, and possibly too small for experimental observation. Consistent

with this, the typical network parameters used in *in vitro* experimental studies of actomyosin contractility are confined to the strong-contraction region (Fig. 5, symbols [35]). Interestingly, these symbols lie between the deformable filaments and the finite motor size contraction domains, suggesting that both mechanisms could play a role in these experiments.

V. DISCUSSION

While the emergence of contractility in strongly organized actomyosin assemblies is well understood, here we consider this process in disordered networks such as those found in nonmuscle cells. Among all possible local contraction models, actin filament deformation (bending or stretching) is most prominent in favoring locally contractile configurations of motors and filaments over locally extensible ones. In this mechanism, filament deformation causes contractility rather than being a mere by-product of it. Local rearrangements due to the motors' finite size could also play a role in *in vitro* experiments. We formulate quantitative predictions of the forces generated by these mechanisms, yielding insights into the influence of the network's microstructure and enabling experimental verifications.

The predicted importance of filament deformation is consistent with *in vitro* studies where the deformation of a reconstituted actomyosin sheet is found to exactly coincide with the amount of deformation of individual filaments, suggesting that filament deformation indeed causes contraction [14]. We also account for the observed inhibition of contractility by excessive cross-linking (\mathcal{D} vanishes for $\xi \rightarrow 0$) [15]. Additionally, the fact that almost parallel filaments dominate contractility in most of the mechanisms studied here is in good agreement with simulations suggesting that filament alignment favors contraction [37]. It would be interesting to extend our results to partially bundled networks—which readily form *in vitro* [38]—knowing that contraction within a bundle also crucially involves filament deformation [12,13]. Note, however, that in the mechanism described here, motors pull on the filaments in both the longitudinal and transverse direction, while in bundles only longitudinal forces are significant. Consequently, motors pulling transverse to a bundle might be much more effective at deforming the actin and thus generating contraction than the motors within, as the latter are deforming the filaments through comparatively ineffective buckling. Finally, we note that *in vitro* parallel bundles of actin filaments contract considerably less than antiparallel bundles [23], in contradiction with a robust prediction of the position-dependent stall force model [24]; this supports our finding that the position-dependent stall force has little effect on contractility. This conclusion could, however, be modified in networks of, e.g., kinesin motors and the stiff filaments microtubules.

Although we find that filament deformations dominate many significant regimes of actomyosin contraction, our focus on local actin deformation could still lead to an underestimation of their effect. Indeed, nonlocal deformations of the network over several mesh sizes could be more favorable than local deformations in heavily cross-linked networks or regimes where motors are larger than the inter-cross-link length. Collective effects could also be of importance, as stress propagation through the elastic filament network could lead to cooperativity between distant motors. We note that our weakly deformed networks approach is only relevant for small motor forces or during the very early stages of larger-scale contraction. Further work is required to analyze strongly deformed or dynamically reorganizing networks and the corresponding synergies between several of the mechanisms described here. On such longer time scales, the microscopic interactions between filaments and motors considered here could furthermore shed light on the self-organization of disordered actomyosin networks into more organized structures [7].

Assessments of the experimental relevance of the mechanisms described here will be facilitated by recent developments in *in vitro* assays [11,14,16,23,30]. Indeed, these now allow precise tuning of the motor and network characteristics as well as detailed monitoring of the network deformations, from which the magnitude of the local force dipole could be inferred. How these considerations apply *in vivo* is a fascinating question, which requires further investigations into alternatives to the paradigm of sarcomerelike contraction.

ACKNOWLEDGMENTS

I thank C. Broedersz, M. Gardel, M. Murrell, and T. Thoresen for countless inspiring discussions, as well as P. Ronceray, A. Roux, and C. Sykes for careful reading of the manuscript. Our group belongs to the CNRS consortium CellTiss. This work was supported by Grants from Université Paris-Sud and CNRS, the University of Chicago FACCTS program, Marie Curie Integration Grant No. PCIG12-GA-2012-334053, and “Investissements d’Avenir” LabEx PALM (ANR-10-LABX-0039-PALM).

-
- [1] A. G. Szent-Györgyi, *The Early History of the Biochemistry of Muscle Contraction*, *J. Gen. Physiol.* **123**, 631 (2004).
 [2] F. S. Fay, K. Fujiwara, D. D. Rees, and K. E. Fogarty, *Distribution of Alpha-Actinin in Single Isolated Smooth Muscle Cells*, *J. Cell Biol.* **96**, 783 (1983).
 [3] L. P. Cramer, M. Siebert, and T. J. Mitchison, *Identification of Novel Graded Polarity Actin Filament Bundles in Locomoting Heart Fibroblasts: Implications for the Generation of Motile Force*, *J. Cell Biol.* **136**, 1287 (1997).

- [4] A. Carvalho, A. Desai, and K. Oegema, *Structural Memory in the Contractile Ring Makes the Duration of Cytokinesis Independent of Cell Size*, *Cell* **137**, 926 (2009).
 [5] O. Medalia, I. Weber, A. S. Frangakis, D. Nicastro, G. Gerisch, and W. Baumeister, *Macromolecular Architecture in Eukaryotic Cells Visualized by Cryoelectron Tomography*, *Science* **298**, 1209 (2002).
 [6] A. B. Verkhovsky, T. M. Svitkina, and G. G. Borisy, *Myosin II Filament Assemblies in the Active Lamella of Fibroblasts: Their Morphogenesis and Role in the Formation of Actin Filament Bundles*, *J. Cell Biol.* **131**, 989 (1995).
 [7] Y. Aratyn-Schaus, P. W. Oakes, and M. L. Gardel, *Dynamic and Structural Signatures of Lamellar Actomyosin Force Generation*, *Mol. Biol. Cell* **22**, 1330 (2011).
 [8] G. Salbreux, G. Charras, and E. Paluch, *Actin Cortex Mechanics and Cellular Morphogenesis*, *Trends Cell Biol.* **22**, 536 (2012).
 [9] J.-F. Joanny and J. Prost, *Active Gels as a Description of the Actin-Myosin Cytoskeleton*, *HFSP J.* **3**, 94 (2009).
 [10] F. C. MacKintosh and A. J. Levine, *Nonequilibrium Mechanics and Dynamics of Motor-Activated Gels*, *Phys. Rev. Lett.* **100**, 018104 (2008).
 [11] T. Thoresen, M. Lenz, and M. L. Gardel, *Reconstitution of Contractile Actomyosin Bundles*, *Biophys. J.* **100**, 2698 (2011).
 [12] M. Lenz, M. L. Gardel, and A. R. Dinner, *Requirements for Contractility in Disordered Cytoskeletal Bundles*, *New J. Phys.* **14**, 033037 (2012).
 [13] M. Lenz, T. Thoresen, M. L. Gardel, and A. R. Dinner, *Contractile Units in Disordered Actomyosin Bundles Arise from F-Actin Buckling*, *Phys. Rev. Lett.* **108**, 238107 (2012).
 [14] M. Murrell and M. L. Gardel, *F-Actin Buckling Coordinates Contractility and Severing in a Biomimetic Actomyosin Cortex*, *Proc. Natl. Acad. Sci. U.S.A.* **109**, 20820 (2012).
 [15] P. M. Bendix, G. H. Koenderink, D. Cuvelier, Z. Dogic, B. N. Koeleman, W. M. Briehner, C. M. Field, L. Mahadevan, and D. A. Weitz, *A Quantitative Analysis of Contractility in Active Cytoskeletal Protein Networks*, *Biophys. J.* **94**, 3126 (2008).
 [16] M. Soares e Silva, M. Depken, B. Stuhmann, M. Korsten, F. C. Mackintosh, and G. H. Koenderink, *Active Multistage Coarsening of Actin Networks Driven by Myosin Motors*, *Proc. Natl. Acad. Sci. U.S.A.* **108**, 9408 (2011).
 [17] D. Mizuno, C. Tardin, C. F. Schmidt, and F. C. Mackintosh, *Nonequilibrium Mechanics of Active Cytoskeletal Networks*, *Science* **315**, 370 (2007).
 [18] N. L. Dasanayake, P. J. Michalski, and A. E. Carlsson, *General Mechanism of Actomyosin Contractility*, *Phys. Rev. Lett.* **107**, 118101 (2011).
 [19] T. B. Liverpool and M. C. Marchetti, *Bridging the Microscopic and the Hydrodynamic in Active Filament Solutions*, *Europhys. Lett.* **69**, 846 (2005).
 [20] More rigorously, \mathcal{D} is the trace of a second-rank force dipole tensor $D_{\mu\nu} = \sum_{i,a} (r_i^a)_\mu (f_i^a)_\nu$. Far-field contractility is characterized by the integrated radial stress exerted on a far away sphere. As a rotationally invariant scalar linear in the f_i^a 's due to the linear elasticity of the rigid medium, this quantity can depend on $D_{\mu\nu}$ only through $\mathcal{D} = D_{\mu\mu}$.

- [21] See Supplemental Material at <http://link.aps.org/supplemental/10.1103/PhysRevX.4.041002> for detailed mathematical derivations.
- [22] T. Sanchez, D. T. N. Chen, S. J. DeCamp, M. Heymann, and Z. Dogic, *Spontaneous Motion in Hierarchically Assembled Active Matter*, *Nature (London)* **491**, 431 (2012).
- [23] A.-C. Reymann, R. Boujemaa-Paterski, J.-L. Martiel, C. Guérin, W. Cao, H. F. Chin, E. M. De La Cruz, M. Théry, and L. Blanchoin, *Actin Network Architecture Can Determine Myosin Motor Activity*, *Science* **336**, 1310 (2012).
- [24] K. Kruse and F. Jülicher, *Actively Contracting Bundles of Polar Filaments*, *Phys. Rev. Lett.* **85**, 1778 (2000).
- [25] K. Kruse and F. Jülicher, *Self-Organization and Mechanical Properties of Active Filament Bundles*, *Phys. Rev. E* **67**, 051913 (2003).
- [26] To understand this decrease in the stall force, consider that the motor has a position-dependent force-velocity relationship. At any given position, motor velocity decreases with increasing opposing force. In the middle of the filament, the velocity in the absence of force is substantial, and correspondingly the stall force is significantly larger than zero. As the barbed end of the filament is approached, the velocity at zero force vanishes, implying that the motor's stall force goes to zero.
- [27] T. Odijk, *Stiff Chains and Filaments under Tension*, *Macromolecules* **28**, 7016 (1995).
- [28] The assumption of entropic elasticity is justified for filaments subjected to forces much smaller than their buckling force, which is implicit in the rigid filament assumption used here.
- [29] T. B. Liverpool, M. C. Marchetti, J.-F. Joanny, and J. Prost, *Mechanical Response of Active Gels*, *Europhys. Lett.* **85**, 18007 (2009).
- [30] T. Thoresen, M. Lenz, and M. L. Gardel, *Thick Filament Regulation of Contractility in Self-Organized Actomyosin Bundles*, *Biophys. J.* **104**, 655 (2013).
- [31] H. Kojima, A. Ishijima, and T. Yanagida, *Direct Measurement of Stiffness of Single Actin Filaments with and without Tropomyosin by In Vitro Nanomanipulation*, *Proc. Natl. Acad. Sci. U.S.A.* **91**, 12962 (1994).
- [32] S. S. Rosenfeld, J. Xing, L.-Q. Chen, and H. Lee Sweeney, *Myosin IIB Is Unconventionally Conventional*, *J. Biol. Chem.* **278**, 27449 (2003).
- [33] G. H. Koenderink, Z. Dogic, F. Nakamura, P. M. Bendix, F. C. MacKintosh, J. H. Hartwig, T. P. Stossel, and D. A. Weitz, *An Active Biopolymer Network Controlled by Molecular Motors*, *Proc. Natl. Acad. Sci. U.S.A.* **106**, 15192 (2009).
- [34] J. Alvarado, M. Sheinman, A. Sharma, F. C. MacKintosh, and G. H. Koenderink, *Molecular Motors Robustly Drive Active Gels to a Critically Connected State*, *Nat. Phys.* **9**, 591 (2013).
- [35] The symbols' coordinates are computed by assimilating ξ to the networks' entanglement length $l_e = 620 \text{ nm}/c^{2/5}$, with the actin concentration c in units of mg/mL [36].
- [36] H. Isambert and A. C. Maggs, *Dynamics and Rheology of Actin Solutions*, *Macromolecules* **29**, 1036 (1996).
- [37] N. L. Dasanayake and A. E. Carlsson, *Stress Generation by Myosin Minifilaments in Actin Bundles*, *Phys. Biol.* **10**, 036006 (2013).
- [38] T. T. Falzone, M. Lenz, D. R. Kovar, and M. L. Gardel, *Assembly Kinetics Determine the Architecture of α -Actinin Crosslinked F-Actin Networks*, *Nat. Commun.* **3**, 861 (2012).